Neuron Article

Cortical Firing and Sleep Homeostasis

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SUMMARY

The need to sleep grows with the duration of wakefulness and dissipates with time spent asleep, a process called sleep homeostasis. What are the consequences of staying awake on brain cells, and why is sleep needed? Surprisingly, we do not know whether the firing of cortical neurons is affected by how long an animal has been awake or asleep. Here, we found that after sustained wakefulness cortical neurons fire at higher frequencies in all behavioral states. During early NREM sleep after sustained wakefulness, periods of population activity (ON) are short, frequent, and associated with synchronous firing, while periods of neuronal silence are long and frequent. After sustained sleep, firing rates and synchrony decrease, while the duration of ON periods increases. Changes in firing patterns in NREM sleep correlate with changes in slow-wave activity, a marker of sleep homeostasis. Thus, the systematic increase of firing during wakefulness is counterbalanced by staying asleep.

INTRODUCTION

During non-rapid eye movement (NREM) sleep the electroencephalogram (EEG) shows characteristic slow waves that can be recorded over the entire cortical surface (Massimini et al., 2004). It is well known that slow wave activity (SWA, the NREM EEG power between 0.5 and 4 Hz) increases after periods of wakefulness and decreases after periods of sleep (Achermann and Borbely, 2003). For example, staying awake from \sim 3 to \sim 24 hr results in progressively higher SWA levels at sleep onset, while naps during the day reduce SWA the following night (Tobler and Borbely, 1986; Vyazovskiy et al., 2006; Werth et al., 1996b). Also, SWA peaks early on during sleep and decreases thereafter along with the decline in sleep pressure (Achermann and Borbely, 2003). There is also evidence for a regional regulation of slow waves (Cajochen et al., 1999; Oleksenko et al., 1992), and recent studies show that cortical areas that have been "used" more during waking show higher SWA relative to less engaged areas (e.g., Huber et al., 2004; Kattler et al., 1994), whereas areas that have been "used" less show reduced SWA (Huber et al., 2006). Thus, at least under acute conditions, sleep SWA can be considered a reliable EEG marker of sleep need and may thus be related to sleep function (Tononi and Cirelli, 2006).

At the cellular level, it is well known that cortical neuronal firing patterns are characteristically different in NREM sleep compared to both wakefulness and REM sleep (Burns et al., 1979; Desiraju, 1972; Hobson and McCarley, 1971; Murata and Kameda, 1963; Noda and Adey, 1970, 1973; Steriade et al., 2001; Verzeano and Negishi, 1960). Intracellular recordings have shown that, during NREM sleep, virtually all cortical neurons engage in the slow (<1 Hz) oscillation, consisting of a depolarized up state, when neurons show sustained firing, and a hyperpolarized down state, characterized by neuronal silence (Amzica and Steriade, 1998; Destexhe et al., 1999; Steriade et al., 1993d, 2001). There is a close temporal relationship between these cellular phenomena and simultaneously recorded slow (or delta) waves, which are defined as surfacenegative EEG events that fall in the SWA frequency range (Amzica and Steriade, 1998; Contreras and Steriade, 1995). Specifically, the surface negativity in the EEG signal (or depth positivity in the local field potential, LFP) corresponds to the down state of cortical neurons as recorded intracellularly and to the suppression of spiking activity as recorded extracellularly, suggesting that EEG or LFP slow waves are a reflection of nearsynchronous transitions between up and down states in large populations of cortical neurons (Burns et al., 1979; Calvet et al., 1973; Contreras and Steriade, 1995; Ji and Wilson, 2007; Luczak et al., 2007; Molle et al., 2006; Mukovski et al., 2007; Murata and Kameda, 1963; Noda and Adey, 1973; Steriade et al., 1993c, 2001).

Thus, (1) sleep EEG slow waves reflect the transition between up and down states of cortical neurons, and (2) EEG SWA is a marker of sleep homeostasis and presumably of sleep need. One might then hypothesize that some aspects of cortical firing may change in relation to sleep pressure. In other words, do cortical neurons fire differently depending on how long the brain has been awake? Surprisingly, this basic question has never been addressed. To fill this gap, we recorded continuously for several days EEG and cortical unit activity in freely behaving rats, during spontaneous sleep/waking cycles as well as after sleep deprivation. We report here that cortical firing patterns do indeed change as a function of sleep homeostasis, in terms of neuronal firing rates, firing synchrony, and distribution of ON and OFF periods.



Figure 1. Cortical Activity in Sleep and Waking

(A and B) Hypnogram, EEG traces from the right barrel cortex and corresponding electromyogram (EMG) in a representative rat during a 2 hr interval of undisturbed baseline starting at light onset (positivity is upward).

(C) Average EEG power spectra in NREM sleep, REM sleep, and waking (mean + SEM, n = 6 rats). Note high values of spectral power in the slow waves range (SWA, 0.5–4.0 Hz; gray bar) in NREM sleep.

(D) Raw multiunit activity (MUA) recorded simultaneously in the same rat from a microwire array placed in the left barrel cortex (six individual channels are shown). Note high tonic firing in waking and REM sleep, and OFF periods in NREM sleep.

(E) Raster plots of spike activity for the same 6 channels shown in (D) (each vertical line is a spike). Note the close temporal relationship between OFF periods and the negative phase of EEG slow waves. Spike sorting was done according to a standard technique (see Experimental Procedures and Figure S1) and the recorded neural population was stable over time (Figure S2).

RESULTS

The Negative Phase of EEG Slow Waves Corresponds to OFF Periods in Cortical Multiunit Activity

The EEG during waking and REM sleep is characterized by theta (6–9 Hz) waves and fast frequencies, while in NREM sleep is dominated by high amplitude and low frequency (0.5–4.0 Hz) slow (delta) waves, which account for most of the power in the EEG spectrum (Figures 1A–1C). Neuronal activity in the barrel cortex, measured extracellularly using microelectrode arrays (see Figures S1 and S2 available online), also changes dramatically between sleep and waking, with periodic total suppression of neuronal firing during NREM sleep (Figures 1D and 1E). We call the periods when all recorded neurons are silent for at least 50 ms OFF periods, as opposed to ON periods, when at least a subset of them shows sustained firing (Figure 1E; average duration, ON periods = 815.5 ± 119.9 ms; OFF periods = 85.8 ± 5.9 ms, n = 6 rats). The OFF periods occur nearly simultaneously with the negative phase of the slow waves on the surface EEG

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(Figures 1C–1E). Thus, they likely correspond to the down state of the slow oscillations as recorded intracellularly. We use the terms "ON" and "OFF" periods, instead of "up" and "down" or "depolarized" and "hyperpolarized" states (Bazhenov et al., 2002; Steriade et al., 1993d, 2001), because the periods of neuronal activity and silence were defined based on the population extracellular activity and not based on changes in membrane potential of individual neurons as measured intracellularly.

Analysis of neuronal activity based on consecutive 4 s epochs revealed a rapid decrease of mean cortical firing rates at the transition from waking to NREM sleep and a gradual increase from NREM to REM sleep (Figure 2A). Overall, periods with high firing rate were more frequent in waking and REM sleep, while periods with low firing rates prevailed in NREM sleep (Figure 2B). Still, firing rates were highly variable within each behavioral state (Figure 2B), likely reflecting different "substates." For example, during active waking many neurons showed periods of silence of up to several seconds, followed by robust firing often in association with specific behaviors, such as exploring or grooming



Figure 2. Effects of Vigilance States on Cortical Neuronal Firing

(A) Time course of mean firing rates during wake-NREM sleep and NREM-REM sleep transitions. Mean values (\pm SEM) represented as % of mean firing rates in NREM sleep (n = 6 rats, 213 neurons). Note that firing rates decrease rapidly after the wake-NREM sleep transition and start increasing ~30 s prior to the onset of REM sleep.

(B) Distribution of 4 s epochs in NREM sleep, REM sleep and waking as a function of mean firing rates (n = 6 rats, 213 neurons). Note that high firing rates can be reached in NREM sleep close to the transition to REM sleep (A).

(C) Distribution of ISIs in NREM sleep and REM sleep represented as % of the corresponding values in waking (mean values \pm SEM, n = 6 rats, 187 neurons).

(data not shown). On average, firing rates were \sim 15%–20% higher during active relative to quiet waking (15.4 ± 0.9 versus 13.0 \pm 0.8 Hz, p < 0.001; quiet waking accounted for <10% of total waking time, consistent with previous reports (Huber et al., 2007b). During NREM sleep, compared to waking and REM sleep, there was a greater proportion of short (<20 ms) interspike intervals (ISIs), likely reflecting high intensity firing during the ON periods, as well as a several fold increase in long ISIs (>100 ms), likely reflecting OFF periods (Figure 2C). Such long ISIs were not only associated with the negativity of EEG slow waves (Figure 1) but also correlated with their amplitude. Thus, high-amplitude slow waves were associated with a more profound suppression of unit activity (Figure 3A), whereas longer OFF periods corresponded to EEG slow waves of higher amplitude (Figure 3B). Since early NREM sleep is characterized by more frequent high-amplitude slow waves (Figure S4), these data suggest that duration and number of the OFF periods could reflect sleep homeostasis.

Number and Duration of ON and OFF Periods Change as a Function of Sleep Pressure

To test this, we compared neuronal activity between early sleep, when sleep pressure and SWA are high, and late sleep, when



Figure 3. EEG Slow Wave Amplitude Is Related to the Duration of the OFF Periods

2 H;

200 ms

↑

(A) The profile of average neuronal firing rates in NREM sleep aligned to the negative peak of the EEG slow wave (the peak is not shown, but is indicated by an arrow). Slow waves were subdivided in three categories based on their amplitude (low: 1%-33%, intermediate: 34%-66%, high: 67%-100%), and the corresponding averages of neuronal activity were computed. Mean values (n = 6 rats). Note that high amplitude slow waves are associated with a larger suppression of neuronal activity.

(B) Average EEG signal, aligned to the onset of OFF periods (arrow). All OFF periods were subdivided into three categories: 20-50 ms, 51-100 ms, and >100 ms, and the corresponding averages of the EEG signal were computed (n = 6 rats). Note that the occurrence of OFF periods is consistently associated with negative waves in the surface EEG. Moreover, longer OFF periods are associated with larger slow waves.

sleep pressure has dissipated and SWA is low (Figure 4A). Visual inspection showed a striking difference: during early sleep, when large slow waves predominate, short ON periods alternated frequently with relatively long OFF periods, whereas in late sleep, when large slow waves are rare, ON periods were longer and only occasionally interrupted by short OFF periods (Figure 4B). A quantitative analysis confirmed that ON periods were initially frequent and short and became less frequent and longer as sleep pressure dissipated (Figure 4C). Both the incidence and duration of the OFF periods decreased in the course of sleep (Figure 4D; the decrease in the OFF periods duration was correlated with longer ON periods; p < 0.001 in all six animals). However, while the magnitude of change in the incidence of the ON and OFF periods was similar, the homeostatic change in duration across the day was more pronounced for the ON periods (Figures 4C and 4D). Notably, changes in the number and duration of ON and OFF periods were highly correlated with the decline in SWA in all animals (Figures 4C and 4D). The correlation was strongest for the longest OFF periods (>128 ms) and for low EEG frequencies (<4 Hz; not shown).

Neuronal Synchrony Changes as a Function of Sleep Pressure

In vivo studies have shown that, during early sleep, EEG slow waves are larger and have steeper slopes than in late sleep (Riedner et al., 2007; Vyazovskiy et al., 2007). Computer simulations reproduced these findings and predicted that steeper slow waves might result from a more synchronous recruitment of individual neurons in population ON periods (Esser et al., 2007). Indeed, we found that in early sleep most individual neurons stopped or resumed firing in near synchrony with the rest of the population (Figure 5A). By contrast, in late sleep, the time

21-50 ms 51-100 ms

> 100 ms



Figure 4. Homeostatic Changes in the Patterns of Neuronal Activity during Sleep

(A) NREM SWA (% of 12 hr baseline) and hypnogram of a 12 hr light period in one representative rat.

(B) EEG and raster plots of neuronal activity in early and late NREM sleep in one representative rat.

(C) Left: changes in incidence and duration of the ON periods during the light phase in one representative rat. Middle: Mean values (±SEM, n = 6 rats) of incidence and duration of the ON periods shown for consecutive 4 hr intervals as percentage of the corresponding mean 12 hr value.

(D) As in (C), but for the OFF periods. Right panels: correlation between SWA (% of 12 hr light period mean) and incidence or duration of the ON and OFF periods in NREM sleep computed for consecutive NREM sleep episodes of the light period in one representative rat (significant correlations were found in all animals).

of entry into ON and OFF periods was much more variable across neurons. To quantify this observation, we computed the latency of the first and last spike of each unit from the onset of population ON or OFF periods, respectively. The synchrony (1/variability) of the latencies decreased by 18% from early to late sleep for ON-OFF transitions, and by 24% for OFF-ON transitions (Figure 5B). We then asked whether changes in neuronal synchronization were related to changes in slow wave slope. Consistent with previous data (Riedner et al., 2007; Vyazovskiy et al., 2007), the slope of surface EEG slow waves was steeper in early sleep compared to late sleep (Figure 5C). Moreover, highly synchronous transitions at the unit level were associated with steep slopes of slow waves, and less synchronous transitions with reduced slopes (Figure 5D). More generally, neuronal synchrony and slow wave slopes were positively correlated (Figure 5E, left panels), whereas the correlation between neuronal synchrony and slow wave amplitudes did not reach significance (not shown). Furthermore, the homeostatic decline of neuronal synchrony at the ON-OFF and OFF-ON transitions correlated with the time course of NREM SWA (Figure 5E, right panels).

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Neuronal Mean Firing Rates Change as a Function of Sleep Pressure

Next, we asked whether neuronal firing rates were also affected by the sleep-wake history. In rats, the longest episodes of wakefulness occur during the night and the longest episodes of sleep during the day. During NREM episodes at night, neuronal firing in the ON periods was intense following sustained wakefulness and much weaker after sustained sleep (Figure 6A, left panels). To quantify this effect, for each animal we selected two NREM sleep episodes-one after the rat had been mostly awake (spontaneously) and another after it had been mostly asleep (50.0 ± 1.6 waking minutes in the preceding hour versus 13.7 ± 2.2 min, respectively). We found that firing rates were significantly higher during the ON periods preceded by prolonged wakefulness compared to those preceded by consolidated sleep periods (Figure 6A, right panel). We then focused on daytime, when rats are predominantly asleep, and compared early with late sleep. As shown in Figure 6B, firing rates during ON periods were consistently higher in early NREM sleep and lower in late NREM sleep. Moreover, changes in firing rates during the ON periods of

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Figure 5. Decreased Synchrony between Individual Neurons in Late Sleep

(A) Raster plots of spike activity in six channels during ON-OFF and OFF-ON transitions in early and late NREM sleep in one representative rat (each vertical bar represents one spike). Vertical dotted lines show the beginning and the end of the single OFF period depicted in the figure, while vertical thick lines indicate, for each neuron within the recorded population (six neurons in this case), the average latency of their last and first spike from the onset of the OFF or ON periods, respectively (to assess their synchrony).

(B) Neuronal synchrony at the ON-OFF and OFF-ON transition measured as 1/standard deviation (in ms) between the latencies of the last and first spike of each neuron from the onset of population OFF and ON transition, respectively (mean values + SEM, 125 neurons, n = 4 rats). Triangles, p < 0.05. (C) Average slopes of the EEG slow waves. Triangles, p < 0.05.

(D) Average surface EEG slow waves aligned to their start point (ON-OFF transition) or their end point (OFF-ON transition). Mean slow waves (SEM, n = 4 rats) are shown for the highest 50% and lowest 50% among all ON-OFF and OFF-ON transitions based on the synchrony between individual units (computed as in B). (E) Left: relationship between neuronal synchrony at ON-OFF or OFF-ON transitions and the corresponding slow wave slopes (% of mean). For each individual recording day (n = 4 rats, 2–5 days/rat) all ON-OFF and OFF-ON transitions were subdivided into five percentiles based on transition synchrony and the corresponding average slow wave slopes were computed. Right: relationship between NREM SWA (0.5–4.0 Hz, % of 12 hr light period mean) and neuronal synchrony at the ON-OFF and OFF-ON transitions of the light period (n = 7 rats, 1–5 days/rat). Lines depict linear regression (Pearson).

NREM sleep correlated positively with changes in NREM SWA (Figure 6C).

Are these changes in firing rate occurring exclusively during NREM sleep, or are they found in all behavioral states? As shown in Figure 6D, during episodes of wakefulness and REM sleep mean firing rates were also higher under high sleep pressure and lower under low sleep pressure. These changes in firing rates were even more remarkable if one considers that rats are highly polyphasic and, especially during the dark period, the spontaneous sleep/wake cycle is fragmented. To further test whether even short wake and sleep episodes lead to changes in neuronal activity, we selected short (5-25 min) continuous wake episodes preceded and followed by sleep, or sleep periods (\leq 60 min) consisting mainly of NREM sleep and preceded and followed by waking, and compared firing rates at the beginning and end of these episodes. We found that the wake-related increase in firing rate was present even after short waking bouts but only if waking was not interrupted by sleep attempts (Figure 6E), suggesting that the quality of waking is an important determinant of neuronal activity during subsequent sleep. At the same time, even short (<15 min) "power naps" led to a significant decrease in firing rates, although the effect was more pronounced after longer sleep periods (Figure 6F).

Though multielectrode arrays are biased toward picking up activity from larger and closest cells, we could detect spikes from all three major neuronal subtypes: putative regular spiking (RS), intrinsically bursting (IB), and fast-spiking (FS) neurons (Figures 7A and S3). Consistent with previous studies (Bartho et al., 2004; McCormick et al., 1985; Steriade et al., 1993c), these three types were subdivided into two groups based on the width of the action potential at 1/2 amplitude (Figures 7B and 7D). The group with narrow spikes (<0.25ms) was characterized by low spike amplitude, higher firing rates and shorter ISIs, and therefore likely includes mostly putative inhibitory neurons (FS cells). The group with broad spikes (>0.25 ms) was characterized instead by high spike amplitude, longer ISIs or spontaneous bursts, overall lower rate of discharge, and likely includes mostly pyramidal excitatory neurons (RS and IB cells). Computing the firing rates for the high and low sleep pressure condition separately for neurons with narrow and broad spikes revealed that they were similarly affected by the sleep/wake history (Figure 7E). Altogether, these findings indicate that cortical mean firing rates change as a function of sleep homeostasis, and they do so across all three behavioral states.



Figure 6. Effects of Sleep/Waking History on Firing Rates

(A) Top left: SWA time course and corresponding hypnogram during an \sim 3 hr interval during the night, centered on an \sim 40 min long waking bout in one representative rat. Bottom left: examples of NREM ON periods before and after consolidated waking. Right: average firing rates within the ON periods (mean values + SEM, 115 neurons, n = 4 rats). Triangle: p < 0.05.

(B) Top left: SWA and corresponding hypnogram during a 6 hr interval starting at light onset in one representative rat. Bottom left: examples of ON periods in early (within the first hour after lights on) and late (\sim 5–6 hr after lights on) sleep. Right: average firing rates within the ON periods in early and late sleep (mean values + SEM, 125 neurons, n = 4 rats). Triangle: p < 0.05. Note that since different data sets contributed to (A) and (B), absolute firing rates values cannot be compared directly.

(C) Relationship between NREM SWA (0.5–4.0 Hz, % of 12 hr light period mean) and neuronal firing rates within the ON periods (% of 12 hr light period mean) computed for the four 3 hr intervals of the light period (n = 7 rats, 1–5 days/rat). Line depicts linear regression (Pearson).

(D) Mean firing rates computed for NREM (including ON and OFF periods, 162 neurons, n = 7 rats), waking (106 neurons, n = 7 rats), and REM (136 neurons, n = 7 rats) sleep in conditions of high and low sleep pressure (mean values + SEM). Triangles: p < 0.05. To compare firing rates during waking, 4 s epochs in high and low sleep pressure condition were equated based on EMG values.

(E) Firing rates within the ON periods in NREM sleep before and after waking episodes lasting 5–25 min (n = 7 rats). All waking episodes were subdivided into those without short sleep attempts (0% sleep) and those containing \sim 1%–10% of NREM sleep. Mean values are shown as % of the mean between the bars (+SEM). Triangle: p < 0.05.

(F) Firing rates in waking before and after sleep periods lasting \leq 60 min and consisting of >70% of NREM sleep. Firing rates are shown separately for short sleep periods <15 min and those longer than 15 min. Mean values are shown as % of the mean between the bars. Triangles: p \leq 0.05.

Increased Sleep Pressure after Sleep Deprivation Is Associated with Changes in Neuronal Activity patterns and Elevated Firing Rates

EEG SWA is a marker of homeostatic sleep pressure, being proportional to the time spent awake, and is largely unaffected by circadian time (Dijk et al., 1987). The changes in neuronal firing patterns and firing rates reported here appear to follow SWA, suggesting that they are homeostatic. To test the homeostatic nature of these effects and experimentally disentangle them from circadian effects, we performed 4 hr of sleep deprivation starting at light onset and measured cortical activity during the subsequent recovery sleep. Sleep deprivation was successful (rats were awake 93% of the time) and, as expected, recovery sleep was associated with elevated SWA compared to the corresponding time of day during baseline (SWA in the first 2 hr interval of recovery: 195.6 ± 16.2 versus 103.2 ± 4.0, % of baseline mean, p < 0.01; Figure 8A). During sleep deprivation, neuronal firing rates showed a progressive increase up to the third hour and then reached a plateau during the fourth hour (Figure 8B). To investigate in more detail which changes in neuronal activity patterns could account for this time course, we quantified the number of long (>50 ms) and short (<20ms) interspike intervals (ISIs) for each individual neuron. Both measures increased progressively, suggesting that while neurons showed increased



Figure 7. Effects of Sleep-Wake History on the Firing Rates of Different Neuronal Subtypes

(A) Interspike intervals (ISIs) distribution for three representative neurons belonging to three major firing phenotypes, putative fast spiking (FS), putative regular spiking (RS), and putative intrinsically bursting (IB). Insets show the shortest ISIs (<20 ms). Note the different scales on the y axes.

(B) Distribution of individual neurons as a function of their spike width at ½ amplitude. All units were subdivided into two categories with narrow spikes and broad spikes (shaded areas).

(C) Average spike waveforms corresponding to the two categories of narrow spike (<0.25 ms) and broad spike (>0.25 ms) units.

(D) Interspike intervals (ISIs) distribution for neurons characterized by short action potential (<0.25 ms, putative inhibitory neurons, 45 neurons, n = 6 rats) and for neurons characterized by long action potential (>0.25 ms, putative excitatory neurons, 38 neurons, n = 6 rats). Mean values ± SEM. Inset highlights the differences between the two neuronal subtypes for the short ISIs (<20 ms).

(E) Mean values (+SEM, n = 6-7 rats/group) of firing rates in NREM sleep, REM sleep and waking computed separately for the narrow spike and broad spike units for high and low sleep pressure conditions. Firing rates are shown as absolute values (top) and as % of the mean between the bars (bottom). Note the different scales on the y axes. Triangles: p < 0.05.

firing as reflected in the short ISIs (>40% increase), this increase was counteracted at the end of sleep deprivation by an increased number of neuronal silent periods (Figure 8B).

After sleep deprivation the duration of ON periods during early recovery sleep was \sim 40% lower than at baseline, whereas both number and duration of OFF periods increased (Figure 8C). In all rats, the decline of these variables toward baseline values in the course of recovery sleep was correlated (p < 0.05) with the decrease in SWA. Despite the much shorter duration of ON periods after sleep deprivation, it was still possible to identify ON periods that were long enough to reveal intense neuronal

firing (Figure 8D). Computation of firing rates during ON periods revealed significantly higher activity (by ~20%) in the first hour after sleep deprivation relative to the corresponding baseline interval (same circadian time; Figure 8E). Neuronal firing rates during early recovery were also affected in waking and REM sleep. Thus, compared to the corresponding baseline interval, in the first hour after sleep deprivation firing rates were increased by 64.4% \pm 21.9% in waking (p < 0.01), and by 33.3% \pm 10.9% in REM sleep (p < 0.01; Figure 8E). Of note, neuronal firing rates in early recovery changed also with respect to pre-sleep deprivation levels: in NREM sleep firing rates increased by



Figure 8. Effects of Sleep Deprivation on Cortical Firing

(A) SWA time course during the light period in baseline and after sleep deprivation (SDep) in one representative rat. Hypnogram from the same animal is shown below.

(B) Time course of neuronal firing rates, and the number of long (>50 ms) and short (<20 ms) interspike intervals (ISIs) in waking during SDep (50 neurons, n = 5 rats). Mean values ± SEM shown as % of the value during the first hour of SDep. Asterisks: p < 0.05. Inset: average firing rates during the first and fourth hour of SDep after equating the 4 s epochs based on EMG values (mean values shown as % of the mean between the two bars + SEM). Triangle: p < 0.05. (C) Number and duration of ON and OFF periods during the first hour of recovery after SDep (Rec) and corresponding time interval during baseline (BSL). Values are mean + SEM (n = 5 rats). Triangles, p < 0.05.

(D) Representative examples of ON periods (boxed) during baseline and recovery sleep in one rat.

(E) Average firing rates within the ON periods during the first 1 hr interval after SDep and the corresponding time interval during baseline in NREM sleep (62 neurons, n = 7 rats) and REM sleep (49 neurons, n = 7 rats). Values are mean \pm SEM.

 $34.0\% \pm 16.3\%$ (p < 0.05), whereas in REM sleep the increase reached $53.7\% \pm 30.3\%$ (p < 0.05). During the initial recovery, NREM sleep was characterized by significantly shorter ON periods as compared to pre-sleep deprivation levels (-37.3% ± 10.3%, p < 0.05), whereas both the duration of the OFF periods and the number of ON and OFF periods were significantly increased (by $32.0\% \pm 11.8\%$, $81.1\% \pm 33.5\%$, and $132.7\% \pm 45.7\%$, respectively, p < 0.05).

DISCUSSION

Since the beginning of unit recordings in vivo, many studies have investigated changes in neuronal firing across behavioral states. Early reports found that, throughout the cerebral cortex, neurons often show a burst-pause pattern in NREM sleep, compared to tonic firing in wakefulness and REM sleep (Burns et al., 1979; Calvet et al., 1973; Desiraju, 1972; Hobson and McCarley, 1971; Murata and Kameda, 1963; Noda and Adey, 1970, 1973; Verzeano and Negishi, 1960). Later, intracellular recordings in vivo showed that during NREM sleep virtually all cortical neurons alternate between a depolarized up state, during which they are spontaneously firing, and a hyperpolarized, silent down state (Steriade et al., 2001). This so-called slow oscillation (<1 Hz) between up and down states occurs more or less synchronously across many neurons (Steriade et al., 1993b), leading to an alternation between periods of multiunit activity

and silence in extracellular recordings (Steriade et al., 2001) and to negative peaks (slow waves) in the sleep EEG. By contrast, wakefulness and REM sleep are characterized by a sustained depolarization and tonic firing and are associated with an absence of EEG slow waves.

So far, however, no study had asked whether cortical neuronal firing patterns change not just depending on whether one is awake or asleep, but rather depending on how long one has been awake or asleep. This question is important because sleep is homeostatically regulated: the longer one has been awake, the more one needs to sleep; conversely, sleep need can only be reduced by having slept (Achermann and Borbely, 2003). This homeostatic regulation is a main reason to think that sleep may serve an essential function for brain cells (Cirelli and Tononi, 2008). Therefore, knowing whether firing patterns change depending on how long the brain has been awake or asleep might shed light on the need for sleep at the cellular level. By recording continuously EEG and cortical unit activity in freely behaving rats, we found that neuronal firing rates, firing synchrony, and distribution of ON and OFF periods do indeed change as a function of sleep homeostasis.

Homeostatic Changes in Neuronal Firing Rates

A key finding of this study is that spontaneous firing rates of cortical neurons increase in a systematic manner with the duration of preceding wakefulness and decrease with the duration of preceding sleep. These changes in firing rate, which were correlated with changes in sleep homeostasis, were evident across wakefulness, NREM sleep, and REM sleep, suggesting an overall shift in neuronal excitability independent of the current behavioral state. It is generally assumed that average firing rates ought to be relatively stable thanks to the tight balance between excitation and inhibition (Haider et al., 2006). For example, it has been shown that a dynamic equilibrium of excitation and inhibition keeps spontaneous network activity within physiological ranges and prevents paroxysmal activity (McCormick and Contreras, 2001). Moreover, the tight balance between excitation and inhibition is maintained during the middle portion of sleep up states, although excitation is briefly uncoupled from inhibition at the up-down and down-up transitions (Haider et al., 2006). Functionally, firing rates should be regulated to avoid (1) increased energy requirements (Attwell and Laughlin, 2001; Laughlin et al., 1998), (2) neurotoxicity (Choi, 1988), and (3) risk of seizures (McCormick and Contreras, 2001). As shown here, however, merely staying awake leads to a progressive increase in firing rates. Consistent with this finding, (1) in mice, cortical metabolic rates increase after a comparable period (4 hr) of wakefulness and decrease with sleep (Vyazovskiy et al., 2008b); (2) in rats, the levels of glutamate in the cortical extrasynaptic space increase progressively during wakefulness and decrease during NREM sleep (Dash et al., 2009); (3) sleep deprivation leads to increased cortical excitability, resulting in lowered threshold for epileptic activity (Badawy et al., 2006; Civardi et al., 2001; Rowan et al., 1982; Scalise et al., 2006). Thus, it would seem that, notwithstanding various mechanisms promoting stable activity levels, staying awake leads to a progressive increase in firing rates of cortical neurons, which is counterbalanced by staying asleep. The changes in firing rates that we observed between the high and low sleep pressure conditions were usually in the range of a few action potentials per neuron per second. While an increase in 1–2 Hz in the firing rate of individual neurons may be modest in relation to specific stimuli or behaviors, a generalized increase of spontaneous firing rates across the entire neuronal population and across all conditions is likely significant as it may raise the already high metabolic costs of brain activity (Laughlin et al., 1998). Specifically, it has been estimated that an increase in activity of just 1 action potential/cortical neuron/s can increase glucose consumption by 21 μ mol/100 g gray matter/min (Attwell and Laughlin, 2001; Sokoloff et al., 1977). This is a substantial change if one considers that the total average glucose consumption in the awake rodent cortex ranges from 107 to 162 μ mol/100 g/min (Sokoloff et al., 1977).

Increased neuronal firing rates after spontaneous waking episodes suggested that a further increase could occur during extended wakefulness, a possibility that we tested by sleep depriving rats for 4 hr starting at light onset. In most neurons, firing rates increased continuously for the first 3 hr and showed no further significant change in the last hour, most likely due to an increased number of neuronal silent periods. These results are consistent with the observation that cortical glutamate levels no longer increase after a few hours of continuous waking in the rat (Dash et al., 2009). We also found that prolonging wakefulness beyond the usual bedtime led to a further increase in firing rates during recovery sleep. By contrast, when animals were allowed to sleep, firing rates decreased progressively in all behavioral states. Since the comparison between sleep deprivation and sleep was done at the same time of day, changes in mean firing rates must be related to the time the animal spent awake or asleep, rather than to differences in circadian phase. Moreover, these data suggest that under physiological conditions sleep may be important, perhaps even necessary, for returning neuronal activity to a lower, sustainable level after periods of wakefulness. Of note, firing rates decreased from the high to the low sleep pressure condition in neurons with both narrow and broad action potentials. Since these two categories are characterized by high and low mean firing rates and likely correspond to inhibitory and excitatory neurons, respectively, the data suggest that the balance between excitation and inhibition is generally maintained at different levels of physiological sleep pressure.

For the interpretation of the present results it is important to assess the stability of unit recordings over several hours or even days, since changes in the number of spikes could be due to a progressive increase in the number of active units rather than to an increased firing of the same units. As shown in Figure S2, however, waveform analysis (Williams et al., 1999) indicates that the shape of the spikes of individual sorted units remained stable over several hours. Furthermore, while the number of recorded units may change progressively over time due to electrode deterioration and other technical factors, we observed several cycles of systematic increases and decreases of firing with time awake and asleep, respectively. These cycles repeated over several sleep-wake episodes in the course of the same day, or even across different days, and were strongly correlated with SWA. Therefore, our results likely cannot be explained by recording instability.

Homeostatic Changes in Neuronal Synchronization and Relationship to EEG Slow Wave Activity

A second finding was that changes in mean firing rate were accompanied by changes in neuronal synchronization during the transition between ON and OFF periods. During sleep episodes following prolonged bouts of wakefulness (early sleep), individual neurons stopped or resumed firing in near synchrony with the rest of the population. By contrast, during sleep episodes following prolonged bouts of NREM sleep (late sleep), the time of entry into ON and OFF periods was much less synchronous.

In turn, changes in synchrony were accompanied by changes in the duration of population ON and OFF periods. During early sleep, when most neurons were active or silent synchronously, ON periods were short and frequent. During late sleep, the periods of activity and inactivity of individual neurons became progressively less synchronized. Indeed, it was apparent that in late sleep some neurons were unresponsive to the population "drive" and remained in the ON mode while the rest of the population was already silent, or remained silent when most other neurons generated spikes. Consequently, population ON periods became longer and less frequently interrupted by population OFF periods. Without chronic intracellular recordings, it is difficult to know whether the duration of the individual neurons' up state also increases with decreasing sleep pressure, though our computer simulations suggest that this may be the case (Esser et al., 2007).

Finally, we found that changes in neuronal synchrony were linked to changes in the EEG slow waves. Specifically, high firing synchrony was associated with steep slopes of simultaneously occurring EEG slow waves, whereas low synchrony was associated with decreased slopes. These observations provide direct in vivo evidence that EEG slow wave slopes are determined by the rate of recruitment and decruitment of cortical neurons into the slow oscillation, as predicted by modeling work (Esser et al., 2007). Furthermore, changes in the number and duration of ON and OFF periods were also correlated with the decrease in SWA in the course of sleep, consistent with the notion that large and steep EEG slow waves should reflect the nearsynchronous transitions between up and down state in large populations of cortical neurons, whereas reduced synchrony should be associated with smaller and shallower waves (Burns et al., 1979; Calvet et al., 1973; Contreras and Steriade, 1995; Ji and Wilson, 2007; Luczak et al., 2007; Molle et al., 2006; Mukovski et al., 2007; Murata and Kameda, 1963; Noda and Adey, 1973; Steriade et al., 1993c, 2001). While slow wave slopes are likely a function of neuronal synchrony, the mechanisms underlying the changes in slow wave amplitude are less clear, and may be related to the duration of the corresponding periods of neuronal silence (e.g., Figure 3). We found that neuronal synchrony had less pronounced effects on the slope and amplitude of the slow wave corresponding to the OFF-ON transition. Such data are consistent with earlier observations that the second segment of the slow wave is characterized by steeper slope and is less affected by sleep-wake history compared to the first segment (Vyazovskiy et al., 2009; Vyazovskiy et al., 2007). Moreover, in anesthetized ferrets, the transition from down to up state occurs at a significantly faster rate and more synchronously than the transition from activity to silence (Haider et al., 2006).

As mentioned in the Introduction, EEG slow waves are a wellestablished marker of sleep need. In general, the longer one has been awake, the higher the amount of SWA in the sleep EEG; conversely, SWA decreases progressively in the course of sleep (Achermann and Borbely, 2003). However, the cellular correlates of slow wave homeostasis are largely unknown. The present results establish that sleep-wake history is reflected at the level of cortical neuronal activity in terms of both firing rates and firing synchrony. Moreover, there is a tight link between changes in neuronal firing on one hand and homeostatic changes of EEG SWA on the other. Thus, our findings suggest that changes in neuronal firing and synchrony may represent a cellular counterpart of the homeostatic regulation of sleep, raising the possibility that they may have physiological significance.

Possible Mechanisms Underlying Homeostatic Changes in Firing Rate and Synchrony

Why would cortical firing rates and neuronal synchrony increase with the duration of wakefulness and decrease with the duration of sleep? Mechanisms that could be responsible for compensatory changes in firing rates include homeostasis of intrinsic excitability, such as changes in intrinsic conductances (van Welie et al., 2004), as well as global synaptic scaling (Turrigiano, 1999). However, the changes in firing rate observed in vivo in our experiments may be too small to trigger the homeostatic changes observed in vitro or after non-physiologic manipulations during development (Desai et al., 2002).

Another possibility is that changes in neuronal firing and synchrony reflect a progressive, net increase in the strength of cortico-cortical connections during wakefulness, followed by a gradual net decrease during sleep (Tononi and Cirelli, 2006), as suggested by some recent studies (Bellina et al., 2008; Gilestro et al., 2009; Rao et al., 2007; Vyazovskiy et al., 2008a). Indeed, in a large-scale model of the corticothalamic system, a net decrease in the efficacy of excitatory cortico-cortical connections can lead, at the cellular level, to (1) a decrease in mean firing rates both when the model was run in sleep mode and in waking mode, (2) a decrease in population synchrony at the ON-OFF and OFF-ON transitions in sleep mode, and (3) a decrease in the number of ON (and OFF) periods and an increase in their duration (Figure S5). At the EEG level, consistent with previous work (Esser et al., 2007), it led to (4) a decrease in the slope of sleep slow waves and (5) a decrease in SWA. Highly synchronous burst firing during the ON periods of early NREM sleep, the alternation of depolarization and hyperpolarization at around 1 Hz, and the neuromodulatory milieu of sleep may help restore synaptic strength (Czarnecki et al., 2007; Lubenov and Siapas, 2008; Seol et al., 2007). On the other hand, these mechanisms could act to some extent also during wakefulness or on a fast time scale that does not require sustained periods of NREM sleep. Moreover, fast-acting processes of synaptic depression and rebound, operating differentially in excitatory and inhibitory synapses (Galarreta and Hestrin, 1998), could help maintain a balance of synaptic strength without requiring sleep-dependent downscaling. Furthermore, it is uncertain whether and how sleep may affect synaptic homeostasis during development, when

sleep is especially abundant. For example, studies in kittens show that the first few hours of sleep immediately following monocular deprivation are important both to maintain the postsynaptic reduction in evoked responses to the deprived eye and to potentiate the response to the nondeprived eye (Aton et al., 2009; Dadvand et al., 2006; Frank et al., 2001). However, whether the overall effect of sleep after monocular sleep deprivation is a net increase, a net decrease, or no change in synaptic strength, remains unclear, and what might happen during development under more physiological conditions remains unknown.

Finally, changes in the levels of arousal-promoting neuromodulators could in principle account for many of our findings (Figure S5). Neuromodulators such as acetylcholine not only can switch the cortex from a sleep to a wake mode of firing (Steriade et al., 1993a) but can affect both intrinsic and synaptic conductances (Gil et al., 1997; Marder and Thirumalai, 2002). Thus, progressive changes in neuromodulatory levels between early and late sleep may alter the firing patterns of cortical neurons, and our previous computer simulations showed that both lowering synaptic strength and increasing levels of arousal-promoting neuromodulators can decrease SWA and the amplitude of slow waves (Esser et al., 2007). It should be mentioned, however, that the current in vivo experiments showed that changes in mean firing rates as a function of sleep-wake history occurred in all behavioral states, in spite of clear-cut differences in neuromodulatory levels between sleep and waking (Jones, 2005).

Changes in firing after learning tasks or electrical stimulation have been investigated in brain regions outside the cortex, such as the hippocampal formation. Hippocampal recordings have revealed only weak, nonsignificant increases in global firing rates during sleep after exposure to a novel environment (Hirase et al., 2001; Kudrimoti et al., 1999) or after electrically-induced LTP (Dragoi et al., 2003) relative to sleep before, suggesting that hippocampal global firing rates are stable across behavioral states and learning conditions (Buzsaki et al., 2007; Karlsson and Frank, 2008; Pavlides and Winson, 1989). However, it is not clear whether the hippocampus is involved in sleep homeostasis, slow oscillations are absent in several hippocampal subregions (Isomura et al., 2006), and the balance of excitation and inhibition may be regulated differently (Buzsaki et al., 2007). Moreover, in most of these studies, the waking experience was short lasting.

In future studies, it will be important to simultaneously record EEG, neural activity, and the levels of extracellular neuromodulators. Moreover, one should investigate whether the homeostatic changes in firing patterns observed in barrel cortex can be generalized to other cortical areas, including prefrontal areas where slow wave homeostasis is most pronounced (Cajochen et al., 1999; Werth et al., 1996a), and in subcortical areas, including the suprachiasmatic nucleus of the hypothalamus that controls the 24 hr distribution of sleep and waking (Deboer et al., 2003). Also, one would want to know how firing patterns change after tasks that activate circumscribed cortical regions, as suggested by the local regulation of sleep SWA (Huber et al., 2004, 2007a; Vyazovskiy and Tobler, 2008; Faraguna et al., 2008). Moreover, the current findings raise the question of whether the overall increase in firing rate observed across waking reflects a generalized phenomenon or is mainly driven by changes in a subset of neurons, for instance those with sustained firing across most of wake and not those with episodic firing, or those with the highest firing rates, or those exhibiting specific firing patterns (tonic or burst firing). It will also be important to clarify whether firing associated with some behavioral activities, perhaps those leading to experience-dependent plasticity and to the release of neuromodulators, contribute more than others to the overall increase in firing rate during the physiological waking period. If the present findings can indeed be generalized to other cortical regions, they would suggest that sleep may serve, among other functions, to maintain neuronal firing rates at a sustainable level.

EXPERIMENTAL PROCEDURES

Surgery and Multiunit Activity Recording

Male WKY rats were implanted in the left barrel cortex (n = 6) or in the frontal cortex (n = 1) with 16-ch (2 × 8) polyimide-insulated tungsten microwire arrays, as detailed in the Supplemental Data. After surgery, all rats were housed individually in transparent Plexiglas cages. Lighting and temperature were kept constant (LD 12:12, light on at 10 a.m., $23^{\circ}C \pm 1^{\circ}C$; food and water available ad libitum and replaced daily at 10 a.m.). About a week was allowed for recovery after surgery, and experiments were started only after the sleep/waking cycle had fully normalized. Animal protocols followed the National Institutes of Health Guide for the Care and Use of Laboratory Animals and were in accordance with institutional guidelines.

Signal Processing and Analysis

Data acquisition and online spike sorting were performed with the Multichannel Neurophysiology Recording and Stimulation System (Tucker-Davis Technologies Inc., TDT), as detailed in the Supplemental Data. Spike data were collected continuously (25 kHz, 300–5 kHz), concomitantly with the local field potentials (LFPs) from the same electrodes (256 Hz) and surface EEG (256 Hz). Amplitude thresholds for online spike detection were set manually based on visual and auditory control and allowed only crossings of spikes with signal-to-noise ratio of at least 2 (Figures S1 and S2). EEG power spectra were computed by a fast Fourier transform (FFT) routine for 4 s epochs (0.25 Hz resolution). For staging, signals were loaded with custom-made Matlab programs using standard TDT routines and subsequently transformed into the EDF (European Data Format) with Neurotraces software. Sleep stages were scored off-line by visual inspection of 4 s epochs (SleepSign, Kissei), where the EEG, LFP, EMG, and spike activity was displayed. Vigilance states could always be determined.

Experimental Design

Recordings were performed continuously for 2–3 weeks starting from day 5 after surgery, when rats appeared normal and their sleep-wake cycle had normalized. A total of 4–7 animals contributed to different experiments and data analyses. To assure that the homeostatic changes were consistent across days, at least one 12 hr light period and one 12 hr dark period (range, 1–7 days) were selected per animal based on signal stability, and analyzed separately. After 2–3 stable baselines, one or two 4 hr sleep deprivation (SDep) experiments were performed in each animal (at least 5 days apart), each followed by an undisturbed recovery period, as detailed in the Supplemental Data. Analysis of the firing rates is detailed in the Supplemental Data.

Data Processing and Analysis and Spike Sorting Details are given in the Supplemental Data.

Histological Verification

Upon completion of the experiments the position of the arrays was verified by histology in all animals. In all cases, the deep row of the array was located within layer V, whereas the superficial row was in layers II–III.

Computer Simulations

The large-scale computational model of the thalamocortical system is similar to the one previously used (Esser et al., 2007) and is described in detail in the

Supplemental Data. Synaptic strength was varied in a subset (33%) of corticocortical AMPA connections. Several levels of strength decrease were tested (details in Supplemental Data). Data shown refer to a 25% decrease, which was found to be sufficient to induce changes in neuronal activity similar to those observed in in vivo experiments.

SUPPLEMENTAL DATA

Supplemental Data include five figures and Supplemental Experimental Procedures and can be found with this article online at http://www.cell.com/neuron/ supplemental/S0896-6273(09)00637-0.

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